

**NORTHRUP EXHIBIT P**

Crit  
Results

M. Allfrey

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Notes (Signal book of several (not all) results photos with this pen (other (on front) was ~~was~~ permanent ink)

- PCR (HIV - MSP) worked well in integrated-heater devices, gel electrophoresis verified product. Some, but minimal primase (esp. due to known fact that device reaction mixture cycled 1-2 times, then at R.T. for  $\frac{1}{2}$  hr & prior to 20 cycles due to need to re-solder connections - new rxn mixture (prim) was added)
- was able to extract  $\sim 100\%$  of aqueous phase with 200  $\mu$ l (set at 30  $\mu$ l) pipette & load 5-6 wells of electrophoresis channel
- (~~①~~) → calculate power consumed in today's experiment compare to batteries

#### Other Discussion

Last Tues w/ Gary Manilla here (Citrus) along w/  
Russ Higuchi, B.B. Watson, Russ's technician, myself we tried homogeneous detection w/ video CCD over 460 thermal cycles

- pulsed Ne - laser (ILC Laser company, Switz) was tried
  - See LLNL Book (notebook) for details

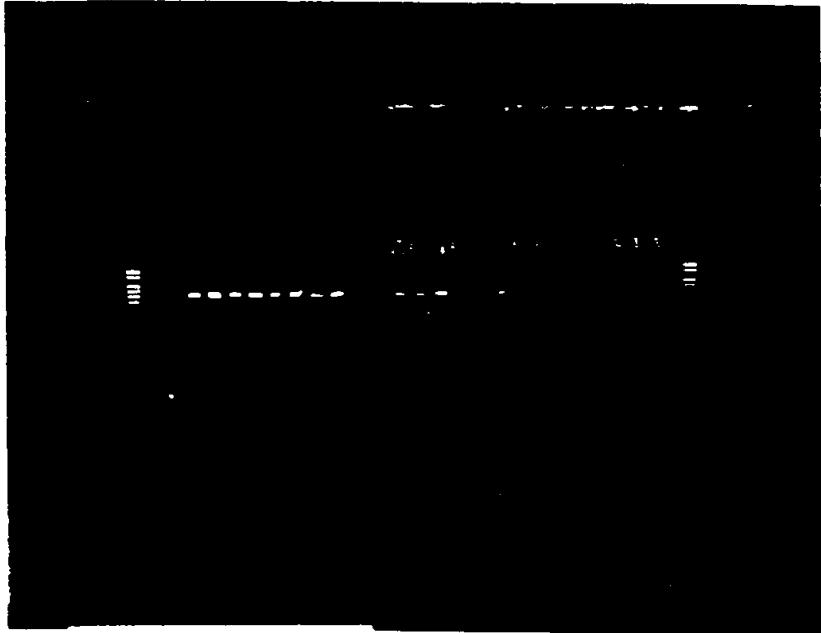
Conf

Results (photos)

Davia PCP results positive

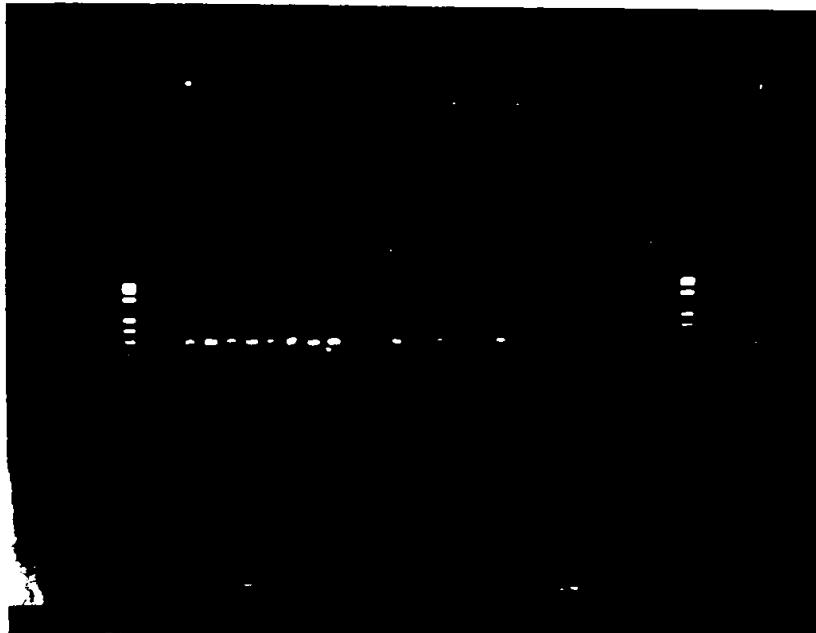
M. All ~~15~~ <sup>15</sup>

elecr. T = 15 min



M. All ~~15~~ <sup>T=2 sec</sup> > 200

elecr. T = 40 min



M. All ~~15~~ <sup>T=1 sec</sup> > 200

taped recip from  
Watson

✓

Rxn	50 $\mu$ l 10X Ref	2
pg		3
B. Watson	50 $\mu$ l 1 mM dATP	
	50 $\mu$ l M13	4
	10 $\mu$ l $10 \times 10 = 100$ pmoles	5
M. Alln/long	400 $\mu$ l	
	10 $\mu$ l 400C	6
	2.5 $\mu$ l $(\times 1.750/\text{each} = 12.5)$	
	$12.5/50/\mu\text{l} = 2.5$	7
	NAQ	
	327.5 $\mu$ l 20	8
	500	

B Cells M. All / <sup>+1</sup>  
J

Tiny new PCR system  
(more Temp. forgiving)

142 bp product target as ss M13 from  
gag-region of HIV

1) Starting target =  $10^8$  copies in 5  $\mu$ l

$T = 96 - 55$   $\downarrow$  16-18 cycles  
(works at 88+)

2) primers

old names: new names  
SK 145 = ph 07  $10 \mu\text{M}/\text{ml}$   
SK 431 = ph 08

Reaction mixture: (500  $\mu$ l)

50  $\mu$ l 10x Buffer w/ mg/ml

" 1 mM dNTPs

" M13 w/ gag region of HIV

10  $\mu$ l =  $10 \times 10 = 100$  pmoles

10  $\mu$ l (same for)  $\frac{\text{ph}07}{\text{ph}08}$  ?

~~500.0  
-172.5  
327.5~~  
2.5  $\mu$ l =  $10 \times 1.25 \mu\text{M}/\mu\text{l}$  12.5

327.5  $\text{H}_2\text{O}$   $\xrightarrow{\text{Tag}}$

500  $\mu$ l Total run volume

M. All / *mf*

(cont.)

- 1) re-use voltage (same device) as  
on March 30 [ie 3.17 V + 98°C]  
at 0.2A

Do only 20 cycles

A) Standards

10, 10, 20, 20, 30, 30, 40, 40

$\sim 15 \mu\text{l}$  oil (1-8)

B) Device 30  $\mu\text{l}$  w  $\sim 90 \mu\text{l}$  oil

1-minute cycles at 3.17V  
20 - 1 minute cycles (A-F) 0.2A

electrophoresis

well-problem

(S) P std #6 1 2 3 4 5 6 7 8 # # A B C D D E F 4 1 4 4  
(F)

- 1c) Had to re-solder device  $\checkmark$  after 2-cycles  
fix time  $\approx 1/2$  hour rxn was  
at room temp

Results - ① formed product in both

stds and in wells

② wells (and 1) std had

less bright primer-dimers

③ device provided  $\sim 6 - 5 \mu\text{l}$  gel

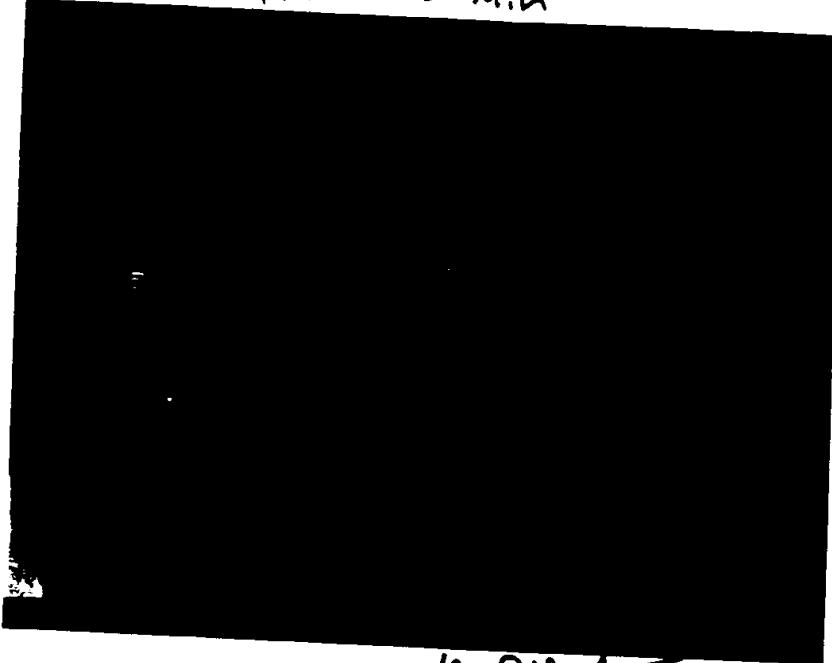
See next  
2 pages:

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Cont results (photos)

M. Allen

electr. Time = 15 min



M. Allen



T = 1 sec 5.6 3200

\*1 loaned to Mila Ching